

**METHOD DEVELOPMENT AND VALIDATION OF SIMULTANEOUS ESTIMATION OF ROSUVASTATIN AND OLMESARTAN IN BULK AND TABLET DOSAGE FORM BY RP-HPLC**Elijabeth.Y*, Ramesh.B¹, K.Dhanalaxmi², Nagarjuna reddy³

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ABSTRACT

A simple, precise, accurate and rapid reverse phase high performance liquid chromatographic new method had been developed for simultaneous estimation of Rosuvastatin and Olmesartan in bulk and tablet dosage form. An Agilent XDB, C18 column having I'd of 150×4.6 mm and 5µm particle size was used. The new method was carried out in gradient program using mobile phase, 0.01M Potassium Dehydrogenate orthophosphate: acetonitrile (55:45 v/v) adjusted to pH-3.2 using dilute ortho phosphoric acid. Flow rate was adjusted to 1.0ml/min and effluents were monitored at 240nm. The retention time obtained for Rosuvastatin and Olmesartan was 2.61 and 5.13min respectively. The calibration curves were linear in the concentration range of 10-30µg/ml for Rosuvastatin and 50-150µg/ml for Olmesartan. The developed method was validated in accordance to ICH guidelines.

Key words: Rosuvastatin, Olmesartan, Acetonitrile, Buffer, RP-HPLC.

INTRODUCTION

Rosuvastatin calcium is chemically [(E)-7[4-(4-fluorophenyl)-6-isopropyl-2-[methyl (methylsulphonyl) amino] pyrimidin-5-yl] (3R, 5S)-3, 5-dihydroxyhept-6-enoic acid] calcium salt. It belongs to a class of drugs called statins, which are employed to lower hypercholesterolemia and related conditions and to prevent cardiovascular diseases. It increases the number of hepatic low density lipoprotein receptors involved in the catabolism of LDL and also inhibits hepatic synthesis of very low density lipoprotein. A detailed survey of analytical literature for Rosuvastatin revealed few methods based on a variety of techniques such as UV-spectrophotometer, high performance thin layer chromatography (HPTLC) and HPLC. Since a HPLC method has many advantages over that of a HPTLC method for quantization, HPLC is often the first choice for developing an analytical method as compared to HPTLC. Till date, none of the reported analytical procedure describes a simple, satisfactory and

validated HPLC method for studying the effect of stress on pharmaceutical dosage forms as well as for assay and determination of content uniformity of Rosuvastatin calcium in tablet dosage forms. Olmesartan medoxomil (OLM) is a prodrug and hydrolyzed to Olmesartan during absorption from the gastrointestinal tract OLM is a selective AT1 subtype angiotensin II receptor antagonist. OLM is described chemically as the (5-methyl-2-oxo-1, 3-dioxol-4-yl) methyl ester of 4-(1-hydroxy-1-methylethyl)-2-propyl-1-[[2-(1H-tetrazol-5-yl) [1, 1-biphenyl]-4-yl] methyl]-1H-imidazole-5-carboxylic acid. Hydrochlorothiazide (HCT), 6-chloro-3, 4-dihydro-2H-1, 2, 4-benzothiadiazine-7-sulphonamide 1, 1-dioxide, one of the oldest and widely used thiazide diuretics. Representative HPLC chromatogram and structures of Olmesartan medoxomil OLM (20 µg mL⁻¹) and hydrochlorothiazide HCT (12.5 µg mL⁻¹). OLM has not yet been officially described in any pharmacopoeia. OLM determination has been reported for single preparations or in combination with other antihypertensive drugs. The

USP describes an RP-HPLC method for the determination of HCT in tablets. Several analytical methods have been reported for the determination of HCT in pharmaceutical formulations including polarography, LC, HPTLC, and spectrofluorometry. A literature survey revealed that analytical methods have not been reported for the determination of OML and HCT in a combined tablet formulation. The present study was therefore aimed to provide such an economically viable RP-LC and HPTLC method.

MATERIALS AND METHOD

Chemicals and solvents: Pure samples of ROSUVA and OLME were obtained respectively from Spectrum pharma research solutions, Hyderabad, India. The commercial pharmaceutical preparation XARB-H containing 20mg and 40mg ROSUVA and OLME respectively (Marketed by Piramal Health care Pvt. Ltd.) were procured from local pharmacy. Acetonitrile, Methanol and water used are of HPLC grade.

Instrumentation: The chromatographic separations were performed using HPLC-Waters alliance (Model-2695) consisting of an inbuilt auto sampler, a column oven and 2996 PDA detector. The data was acquired through Empower-2-software. The column used was Alliance C18 (150×4.6mm I'd, 5µm particle size). Meltronics sonicator was used for enhancing dissolution of the compounds. Dig sun pH meter was used for adjusting the pH of buffer solution. All weighing was done on sarotorious balance (model AE-160).

Chromatographic condition of method: An Agilent XDB, C18 column having I'd of 150×4.6 mm and 5µm particle size was used. At ambient temperature. 1.36 g of Potassium dihydrogen orthophosphate was weighed and 1000ml of Milli-Q water was added to it. The mobile phase was considered buffer: acetonitrile. PH was adjusted to 3.2(0.02) with Ortho phosphoric acid and was filtered through 0.45µm PVDF membrane filter disc and was degassed. Flow rate was maintained at 1ml/min. The elution was observed at 240nm. Mobile phase was 55:45 (buffer: Actonitrile). Injection volume and runtime were 20 µl and 10 mins respectively. In the ratio 55:45 retention time for Olmesartan and Rosuvastatin were observed to be 2.61 and 5.13 min respectively. The two peaks were well resolved with good peak shape and symmetry was obtained. Hence this method was finalized for the simultaneous estimation of Olmesartan and Rosuvastatin

Preparation of buffer solution: Accurately weighed 1.36 gm of Potassium Dehydrogenate orthophosphate (0.02M) was transferred into 1000ml volumetric flask. Add about 900ml of Milli-Q water and Sonicate to dissolve, then make up to 1000ml and then pH was adjusted to 3.4 using dilute Ortho phosphoric acid.

Preparation of mobile phase: The prepared phosphate buffer solution and acetonitrile was used as mobile phase in ratio 55:45.

Preparation of standard stock solutions: Accurately Weighed and transferred 20mg of Olmesartan and 40mg of Rosuvastatin and working Standards into a 10 ml clean dry volumetric flask, add 7ml of diluents, sonicated for 5 minutes and make up to the final volume with diluents.(standard stock). This was used for calibration purpose of both the drugs.

Preparation of sample solution: 5 tablets were weighed and calculate the average weight of each tablet then the weight equivalent to 5 tablets was transferred into a 100 mL volumetric flask, 80mL of diluents added and sonicated for 25 min, further the volume made up with diluents and filtered. From the filtered solution 0.2ml was pipette out into a 10 ml volumetric flask and made up to 10ml with diluents.

METHOD VALIDATION

The developed method was validated as per the ICH (International Conference on Harmonization) guidelines with respect to System suitability, Precision, Specificity, Linearity, Accuracy, Limit of detection and Limit of quantification.

Linearity: Aliquots of 0.25, 0.75, 1.0, 1.25, and 1.5 ml were taken from stock solution of concentration 1000µg/ml OLME and 500µg/ml ROSUVA, and then diluted up to mark with methanol. Such that the final concentrations were in the range 10-30µg/ml for OLME and 50-150µg/ml for ROSUVA. Volume of 10µl of each sample was injected in five times for each concentration level and calibration curve was constructed by plotting the peak area versus drug concentration. The observations and calibration curve were shown in Table 1 and Fig. 2, 3.

Estimation of Olmesartan and Rosuvastatin: Accurately weighed powder equivalent to 20mg of OLME and 40mg of ROSUVASTATIN was transferred into 10ml volumetric flask and made up to the mark with diluents methanol to obtain solution of OLME (20mg/ml) and ROSUVA (40mg/ml). From this each solution 0.2ml and 0.4ml was transferred to 10ml volumetric flask and made up to

the mark with diluents methanol to obtain solution of OLME (20µg/ml) and ROSUVA (40µg/ml). The results were shown in Table-2. The chromatograms were shown in Fig-4, 5.

Accuracy: Accuracy of the method was done by recovery study. Sample solutions were prepared by spiking at about 50%, 100%, and 150% of specification limit to placebo and analyzed by the proposed HPLC method. Results are shown in Table-4.

Specificity: The specificity of the method was performed by injecting blank solution(without any sample) and then a drug solution of 20µl injected into the column, under optimized chromatographic conditions, to demonstrate the separation of both OLME and ROSUVA from any of the impurities, if present. As there was no interference of impurities and also no change in the retention time, the method was found to be Specific.

Limit of detection (LOD) and Limit of quantification (LOQ): The parameters LOD and LOQ were determined on the basis of response and slope of the regression equation. The linearity for OLME and ROSUVA was performed from 10-30µg/ml and 2.5-18.75µg/ml respectively.

System precision: Precision is the measure of closeness of the data values to each other for a number of measurements under the same analytical conditions. Standard solution of OLME (20µg/ml) and ROSUVA (40µg/ml) were prepared as per test method and injected for 3 times. Results are shown in Table-4.

Method precision: Three samples were prepared and analyzed as per the test method on same day and three different days and calculated the % RSD for assay of five preparations. Results were shown in Table- 5.

Robustness: Robustness studies were carried out by variations in flow rate, mobile phase compositions and temperature. It was observed that the small changes in these operational parameters did not lead to changes of retention time of the peak interest. The degree of reproducibility of the results proven that the method is robust.

System suitability test: The system suitability was determined by making six replicate injections from

Table-1: Linearity

s.no	Concentration of OLME in µg/ml	OLME area	Concentration of ROSUV in µg/ml	ROSUVA area
1	10	1192845	20	1405952
2	15	1850914	30	1961437
3	20	2480230	40	2635705
4	25	3116334	50	3295767
5	30	3683312	60	3913594

freshly prepared standard solutions. The observed RSD values were well within usually accepted limits ($\leq 2\%$). Theoretical plates, tailing factor, resolution between OLME and ROSUVA were determined. The results are all within acceptable limits summarized in Table-6.

RESULTS AND DISCUSSION

The new method was developed by Olmesartan and Rosuvastatin. Nature of sample, its molecular weight and solubility decides the proper selection of stationary phase. The drugs OLME and ROSUVA were preferably analyzed by reverse phase chromatography and accordingly C₁₈ column was selected. The elution of the compounds from column was influenced by polar mobile phase. The ratio of phosphate buffer to Acetonitrile was optimized to (55:45) to give well resolved and good symmetrical peaks with short run time. The retention time of OLME and ROSUVA were found to be 2.611 & 5.138 min respectively. The linearity of the method was statistically confirmed. RSD values for accuracy and precision studies obtained were less than 2% which revealed that developed method was accurate and precise. The system suitability parameters were given in table-5. The analytical recovery at five different concentrations of OLME and ROSUVA was determined and the recovery results were in the range of 10-30µg/ml. Therefore proposed validated method was successfully applied to determine OLM and ROSUVA in tablet dosage form.

CONCLUSION

The new developed method is accurate, simple, rapid and selective for the simultaneous estimation of OLME and ROSUVA in Tablet dosage form. The retention time of OLME and ROSUVA were found to be 2.611 & 5.138 min respectively. The excipients of the commercial sample analyzed did not interfere in the analysis, which proved the specificity of the method for these drugs. The sample preparation is simple, the analysis time is short and the elution is by gradient method. Hence the proposed method can be conveniently adopted for the future quality control analysis in the combined formulation.

Table-2: OLME and ROSUVA assay:

Drug	Label claim mg/tab	Amount found mg/tab	Label claim(%)	S.D*	% R.S.D
OLME	20	19.90	99.90	22443.53	0.84574
ROSUV	40	39.49	99.93	2948.472	0.101878

Table-3: Accuracy

% linearity level	No. of times	% recovery	%mean recovery \pm S.D	% RSD
50	1	100.3527	100.52 \pm 0.166744	0.165861
	2	100.5608		
	3	100.6825		
100	1	98.3923	99.7905 \pm 1.232173	1.234671
	2	100.6925		
	3	100.3083		
150S	1	100.262	99.6064 \pm 0.791952	0.794119
	2	98.72363		
	3	99.81927		

Table-4: System precession:

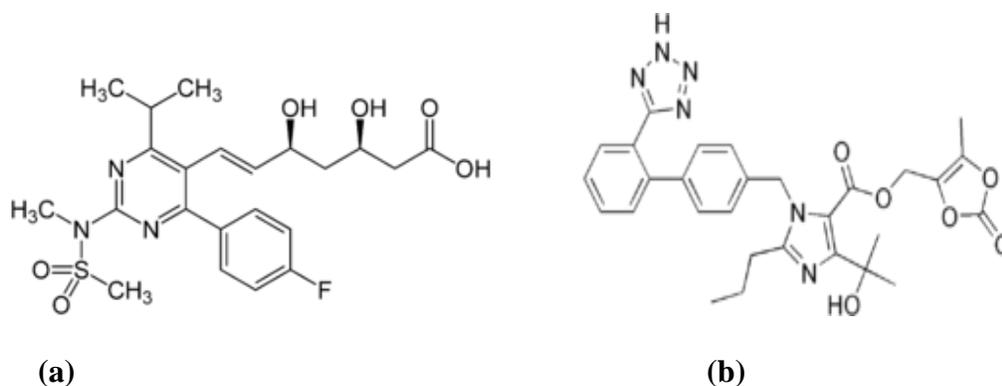
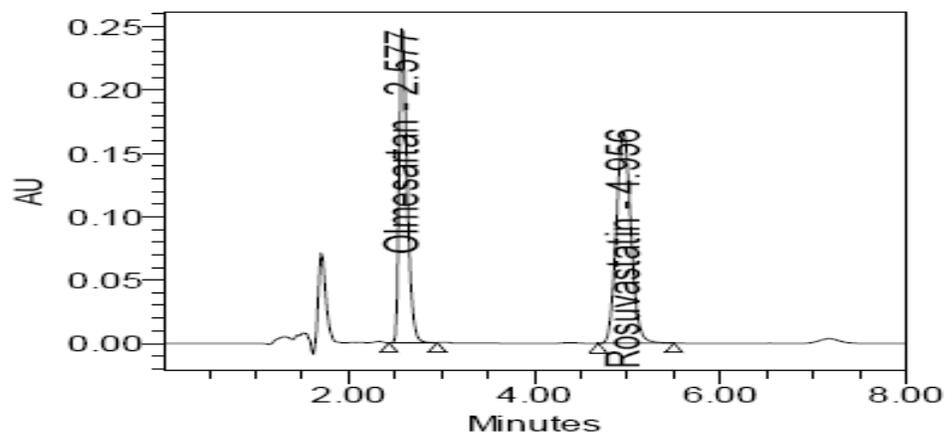
INJECTIONS	AREAS(OLME)	AREAS(ROSUVA)
1	2625657	2897886
2	2634276	2897032
3	2646601	2892929
4	2673054	2894023
5	2658654	2893015
6	2683789	2889859
AVG	2653672	2894124
S.D	22443.53	2948.472
%R.S.D	0.845754	0.101878

Table-5: Method precession:

Drug	% Assay	Mean	S.D	% R.S.D
OLME	98.21623	98.01542	0.250539	0.255612
	98.04171			
	97.57332			
	98.08809			
	97.91057			
	98.26261			
ROSUVA	101.2046	100.2046	1.296163	1.295953
	100.14993			
	98.45324			
	100.9049			
	101.504			
	99.12103			

Table-6: characteristics of HPLC method:

Drug	Parameters defined	Obtained value
Olmesartan	Linearity range ($\mu\text{g/ml}$)	30-225 $\mu\text{g/ml}$
	Slope	123961
	Intercept	12077
	Regression coefficient(r^2)	0.999
	LOD ($\mu\text{g/ml}$)	1.86
	LOQ ($\mu\text{g/ml}$)	5.64
	Tailing factor	1.47
	Plate count	3993
Rosuvastatin	Linearity range ($\mu\text{g/ml}$)	2.5-18.75 $\mu\text{g/ml}$
	Slope	64963
	Intercept	36659
	Regression coefficient(r^2)	0.999
	LOD ($\mu\text{g/ml}$)	3.21
	LOQ ($\mu\text{g/ml}$)	9.74
	Tailing factor	1.10
	Plate count	4854

**Fig. 1. Structures of Rosuvastatin (a) and Olmesartan (b)****Fig.2: chromatogram showing retention time of OLME and ROSUVA:**

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